



Workflows for automated analysis of <u>mouse</u> brain imaging data

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Objective

An efficient way to integrate imaging data sets with variable image resolution and image content is key for cross-modality correlations, e.g., structural to functional or in vivo biomarker to ex vivo histology¹⁻³. We present our atlas-based approach in which all contributing images are accurately aligned through image registration algorithms with the Allen Mouse Brain Atlas (CCF v3)⁴. The atlas resource facilitates the analysis of macroscopic to microscopic features across subjects and integration of viral tracing and gene expression data.



- Individual selection of ROIs somewhere on the image
- Manual delineation of the anatomical regions based on the visible brain structure

Novel approach

We have created automated workflows for the specific needs of cross-modality mouse brain experiments:





- *AIDAdb:* cloud-based relational database⁶
- AIDAqc: MRI quality control
- AIDAmri: automated processing of MRI data⁷
- AIDAhisto: whole slice cell counting⁸
- AIDAconnect: graph theory algorithms for fMRI/DTI data⁹





Workflow histol	ogy		0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
Data aquisition	Registration	Counting	(c) MR8 Textons
Possible input: Immunostainings, confocal and conventional fluorescence microscopy, tif format	Landmark-based using Slicer3D or ImageJ (plugin landmark correspondence)	 Classification of cellular structures with a set of filters Use the segmented cell nuclei to correct the cell counting 	
Input		Result	

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Applications

• § Stroke lesion location and size determine functional deficits and recovery potential¹⁰

• ^{*} Lesion mapping and region-wise analysis of cellular features facilitate the correlation between in vivo and ex vivo data, but also allow to monitor the individual development of the lesion and hence help to answer the question, which brain areas are most relevant for behavioral deficit and recovery of function GFAP/Vim

MRI incidence maps and detailed anatomical lesion mapping **Atlas-based cell counting results** Functional/structural network changes after stroke Atlas Connectivity rs-fMRI Preprocessing rimary motor area Registration Matrix econdary motor area **BOLD-Signa** Retrosplenial area rimary somatosensory area, barrel field imary somatosensory area, lower limb mary somatosensory area, upper limb rimary_somatosensory_area,_mouth **Diffusion MRI** rimary somatosensory area, nose Primary somatosensory area, trunk Primary_somatosensory_area,_unassigned





References

- Hawrylycz et al. Plos Comput. Biology, 2011 https://doi.org/10.1371/journal.pcbi.1001065
- **2.** Hess et al. Mol Imaging Biol, 2018 <u>https://doi.org/10.1007/s11307-018-1259-y</u>,
- Aswendt et al. Mol Imaging Biol, 2017 <u>https://doi.org/10.1007/s11307-016-0988-z</u>, 3.
- Wang et al., Cell, 2020, <u>https://doi.org/10.1016/j.cell.2020.04.007</u>
- 5. Hammelrath et al. NeuroImage, 2016, https://doi.org/10.1016/j.neuroimage.2015.10.009.
- 6. Pallast et al., Oxford Database, 2018, https://doi.org/10.1093/database/bay124
- 7. Pallast et al. Front. Neuroinf., 2019, DOI: <u>https://doi.org/10.3389/fninf.2019.00042</u>,
- Pallast et al. J Neurosci Methods 2019 https://doi.org/10.1016/j.jneumeth.2019.108394,
- Scharwächter et al., Neuroimage, 2022 https://doi.org/10.1016/j.neuroimage.2022.119110,
- 10. Aswendt et al. Transl Stroke Res. 2021 https://doi.org/10.1093/database/bay124.

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